



Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.

Coagulopathy, thromboembolic complications, and the use of heparin in COVID-19 pneumonia



Luca Costanzo, MD,^a Francesco Paolo Palumbo, MD,^a Giorgio Ardita, MD,^a Pier Luigi Antignani, MD,^b Enrico Arosio, MD,^c and Giacomo Failla, MD,^a under the auspices of the Italian Society for Vascular Investigation and the Italian Society of Vascular Medicine, Catania, Rome, and Verona, Italy

ABSTRACT

The SARS-CoV-2 (COVID-19) is causing a pandemic and potentially fatal disease of global public health concern. Viral infections are known to be associated with coagulation impairment; thus, thrombosis, hemorrhage, or both may occur. Understanding the pathophysiologic mechanisms underlying the development of coagulation disorders during viral infection is essential for the development of therapeutic strategies. Coagulopathy in COVID-19 infection is emerging as a precipitant factor for severe respiratory complications and death. An increase in coagulation markers, such as fibrinogen and D-dimer, has been found in severe COVID-19 cases. Heparin, clinically used as an anticoagulant, also has anti-inflammatory properties, including binding of inflammatory cytokines, inhibition of neutrophil chemotaxis, and protection of endothelial cells, and a potential antiviral effect. We hypothesized that low-molecular-weight heparin may attenuate cytokine storm in COVID-19 patients; therefore, low-molecular-weight heparin could be a valid adjunctive therapeutic drug for the treatment of COVID-19 pneumopathy. In this paper, we review potential mechanisms involved in coagulation impairment after viral infection and the possible role of heparin in the treatment of COVID-19 patients. (*J Vasc Surg: Venous and Lym Dis* 2020;8:711-6.)

Keywords: COVID-19; Pneumonia; Thrombosis; Coagulopathy; D-dimer; Low-molecular-weight heparin

Recent studies have shown that coagulopathy can occur during COVID-19 disease. Thromboembolic manifestations such as pulmonary embolism¹⁻³ and disseminated intravascular coagulation (DIC)⁴ have been reported and have resulted in poor prognosis for the patient.

Our hypothesis is that heparin anticoagulation can be useful in COVID-19. Support of this comes from published mechanisms of coagulation impairment after viral infection. The purpose of this publication is to present this rationale.

THE INTERACTIONS BETWEEN INFLAMMATION AND COAGULATION

A correlation between inflammation and coagulation has been widely demonstrated. Inflammation can lead to altered coagulation, with a consequent imbalance

between the procoagulant and anticoagulant states.⁵ Several inflammatory cytokines, such as interleukin 6, interleukin 8, and tumor necrosis factor α , promote a procoagulant state through the expression of tissue factor by a mechanism that includes the activation of endothelial cells, platelets, and leukocytes.⁶ Endothelial cell activation is characterized by the endothelial expression of cell surface adhesion molecules, such as vascular cell adhesion molecule 1, intercellular adhesion molecule 1, and E-selectin.⁷ The platelets are primarily associated with the initiation of coagulation cascades. Platelet activation is stimulated by bound platelet secretion products and local prothrombotic factors. Briefly, after activation, platelets change shape and release the contents of their granules, which contain several molecules, such as inflammatory mediators, hemostatic factors, angiogenic/antiangiogenic factors, growth factors, and protease.⁸ During leukocyte activation, the synthesis and release of numerous inflammatory molecules, such as leukotrienes, prostaglandins, bradykinin, free oxygen radicals, and cytokines, occur.⁹ Therefore, interaction between endothelial cells, platelets, and leukocytes is a crucial element in the immune response after inflammatory stimuli.

Furthermore, an increased release of histones and nucleosomes (DNA + histones), toxic elements to the endothelium, has been shown in sepsis and other inflammatory conditions. In contrast, activated protein C inactivates histones, protecting the endothelium.¹⁰ In response to the infection, extracellular DNA fibers extruded by neutrophils (neutrophil extracellular traps

From the Angiology Unit, San Marco Hospital, Department of Cardiovascular Disease, A.O.U. "Policlinico-Vittorio Emanuele," University of Catania, Catania^a; the Vascular Center, Nuova Villa Claudia, Rome^b; and the Department of Medicine, University of Verona, Verona.^c

Author conflict of interest: none.

Correspondence: Luca Costanzo, MD, San Marco Hospital, Department of Cardiovascular Disease, A.O.U. "Policlinico-Vittorio Emanuele", University of Catania, Viale Carlo Azeglio Ciampi, 95121 Catania, Italy (e-mail: lucacost84@gmail.com).

The editors and reviewers of this article have no relevant financial relationships to disclose per the Journal policy that requires reviewers to decline review of any manuscript for which they may have a conflict of interest.

2213-333X

Copyright © 2020 by the Society for Vascular Surgery. Published by Elsevier Inc.

<https://doi.org/10.1016/j.jvs.2020.05.018>

[NETs]) are produced to allow neutrophils to trap and to kill external microorganisms. NETs stimulate the formation and deposition of fibrin to trap microorganisms and to control infection.¹¹ NETs also cause platelet adhesion, and a link with deep venous thrombosis occurrence has been shown in some experimental models.¹²

D-dimer is an epitope resulting from the plasmin degradation of cross-linked fibrin. D-dimer is elevated in several conditions, such as thrombosis, DIC, and inflammation.¹³ Notably, D-dimer can promote the development of inflammation by activating neutrophils and monocytes, inducing the secretion of some inflammatory cytokines.^{14,15}

THROMBOCYTOPENIA AND VIRAL INFECTION

In several viral infections, the impairment of platelet function, decrease in production, or platelet destruction has been documented. Thrombocytopenia can occur in both hemorrhagic and nonhemorrhagic viral infections, and it is frequently caused by autoimmune antibodies against platelets. Other mechanisms include increased platelet adhesion and activation, resulting in platelet consumption and bone marrow infection that directly affects megakaryocytes and thus platelet production.⁵ In SARS-CoV infection, thrombocytopenia caused by autoantibodies, high levels of von Willebrand factor in the blood,¹⁶ and activation of the coagulation cascade with final generation of fibrin have been described.¹⁷⁻²⁰

In COVID-19 disease, thrombocytopenia at hospital admission does not seem to be a consistent factor; in a study of 41 patients, it was found in only 2 patients.²¹ However, in a recent meta-analysis of nine studies including nearly 400 patients, the platelet count was significantly lower in patients with more severe COVID-19; also, thrombocytopenia was associated with more than fivefold increased risk of severe COVID-19 disease.²²

MARKERS OF IMPAIRED COAGULATION IN VIRAL INFECTION

A procoagulative state can be evidenced through an increase in the levels of coagulation proteins. Increased levels of fibrinogen, D-dimer, thrombin-antithrombin complexes, plasmin- α_2 -antiplasmin complexes, and thrombomodulin have been found in respiratory tract infections, influenza, and SARS-CoV infection. Furthermore, an increased level of the plasminogen 1 activator inhibitor, suggestive of impaired fibrinolysis, was also reported.⁵

Alterations in laboratory parameters compatible with DIC criteria have been described by Tang et al⁴ in 15 deaths for COVID-19. DIC develops as a result of monocyte and endothelial cell activation due to cytokine release after injury, with expression of tissue factor and secretion of von Willebrand factor. The final result is the circulation of free thrombin that can activate platelets

and stimulate fibrinolysis with elevated levels of D-dimer and fibrinogen degradation products. Therefore, in the final stage of disease, high levels of D-dimer and products of the degradation of fibrinogen have been reported; markedly elevated D-dimer levels also correlated with more severe disease and mortality.^{4,23,24}

VIRAL INFECTION AND COAGULOPATHY

Either bleeding or thrombosis has been described as a complication in several viral infections. An exaggerated response to infection can become complicated up to DIC with the formation of microvascular thrombi in various organs.²⁵ Respiratory tract infections increase the risk of deep venous thrombosis and pulmonary embolism.²⁶ In the H1N1 influenza epidemic, both thrombotic and hemorrhagic complications, such as deep venous thrombosis, pulmonary embolism hemorrhage with hemoptysis, hematemesis, petechial rash, and a case of diffuse petechial cerebral hemorrhage, have been reported.⁵ The occurrence of DIC, pulmonary hemorrhage, and thrombocytopenia has been reported in avian influenza (H5N1) in several patients.²⁷ In the severe acute respiratory syndrome induced by coronavirus, vascular endothelial damage in small and medium-sized pulmonary vessels, DIC, deep venous thrombosis, and pulmonary thromboembolism have been described.¹⁸⁻²⁰

Tang et al⁴ reported a consumption coagulopathy in advanced stages of COVID-19 pneumopathy with elevated levels of D-dimer and fibrinogen degradation products. Extremely increased D-dimer results from plasmin-associated hyperactive fibrinolysis, and plasminogen/plasmin system involvement in COVID-19 disease has been investigated in a recent paper.²⁸ Enhanced susceptibility to SARS-CoV-2 infection and fatality have been reported in patients with pre-existing comorbidities including hypertension, diabetes, coronary heart disease, cerebrovascular disease, chronic obstructive pulmonary disease, and chronic renal disease.²³ Elevated plasminogen/plasmin is common in people with several underlying medical conditions, and plasmin has been found to enhance the virulence and infectivity of SARS-CoV-2 virus by cleaving its spike proteins.²⁸

Older age has been reported as a poor prognosticator for COVID-19 survival,²⁴ and many factors could be related. First, several diseases coexist in the elderly, predisposing to endothelial dysfunction and coagulation impairment, as discussed before. Second, immune response is different between young and older patients, and it is known that type I interferon responses in men decrease with age.²⁹ Exacerbated innate host response to SARS-CoV in aged nonhuman primates has been demonstrated with an increase in differential expression of genes associated with inflammation, whereas expression of type I interferon β was reduced.³⁰ Also, a deficiency in control of viral

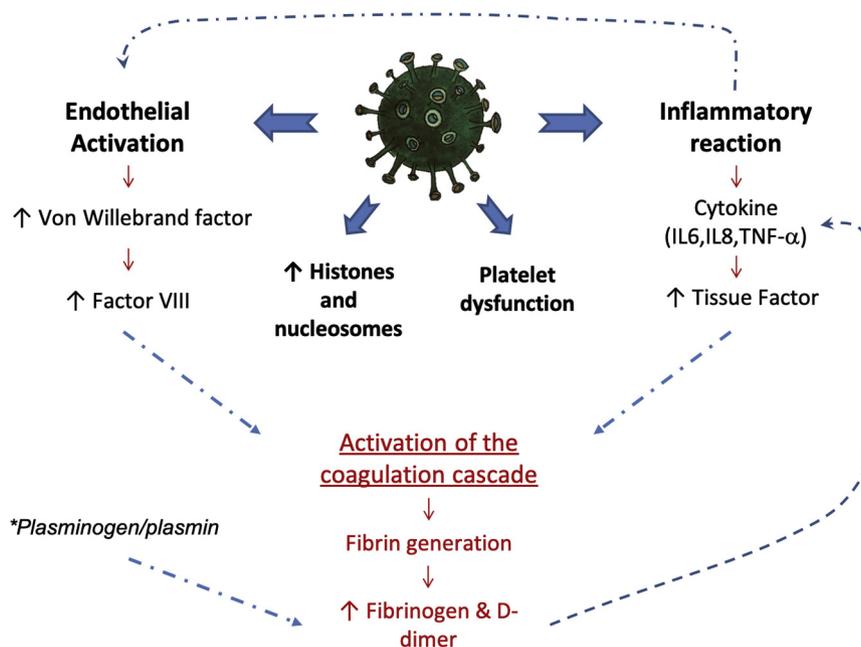


Fig 1. Mechanisms of coagulation impairment in COVID-19 infection. *Elevated levels of plasminogen/plasmin are found in several medical conditions, including hypertension, diabetes, cardiovascular disease, cerebrovascular disease, and chronic renal disease. *IL6*, Interleukin 6; *IL8*, interleukin 8; *TNF-α*, tumor necrosis factor α .

replication could be hypothesized as age-dependent defects in T-cell and B-cell function have been described.³¹

Potential mechanisms of impaired coagulation in COVID-19 infection are summarized in Fig 1.

HISTOPATHOLOGIC FINDINGS

Fibrin accumulation in the lung is a hallmark of acute lung injury and acute respiratory distress syndrome. Fibrin clots in the alveoli have been described in SARS-CoV infection in humans and mice. This coagulation response is hypothesized to protect the host by sealing the alveoli, preventing edema and alveolar hemorrhages but limiting the exchange of oxygen.³² The first published autopsy report of deaths due to severe COVID-19 infection revealed diffuse alveolar damage with fibrin exudation in alveoli, modest infiltration of immune cells in vessel walls, and hyaline thrombi in small vessels and also in other organs and tissues.³³ Similar findings have recently been reported by American pathologists who described a thrombotic microangiopathy in lung associated with foci of alveolar hemorrhage.³⁴

Moreover, a generalized thrombotic microvascular injury has been documented by Magro et al³⁵ in a series of lung and skin biopsies. Apart from the histologic changes described by the other authors, they found deposits of terminal complement components in the microvasculature, consistent with sustained, systemic activation of complement pathways. Therefore, in a subset of severe COVID-19 infections, a vasculitis-like syndrome with an associated procoagulant state can occur.

RATIONALE FOR THE USE OF HEPARINS IN COVID-19 INFECTION

Heparins are anticoagulant drugs currently used for the prophylaxis and therapy of venous thromboembolism and are classified according to their molecular weight.³⁶ Heparin reversibly binds antithrombin III and amplifies its subsequent inhibitory effect on activated factor X and thrombin (factor Xa).^{37,38} Only unfractionated heparin containing at least 18 saccharide sequences can influence the action of antithrombin on thrombin; however, unfractionated heparin fragments of any length containing a unique pentasaccharide sequence can inhibit the action of factor Xa.^{39,40} This feature has been exploited by pharmacologic research for the development of low-molecular-weight heparins (LMWHs).⁴¹

Fondaparinux, a synthetic analogue of the pentasaccharide sequence, has a longer half-life than LMWH and does not interact with platelets.⁴² Fondaparinux binds reversibly to antithrombin to produce an irreversible conformational change that enhances its reactivity with factor Xa. This results in inhibition and depletion of factor Xa, which in turn inhibits thrombin generation in the coagulating signal transduction pathway. Notably, this molecule is characterized by therapeutic coverage for 24 hours and noninterference with platelets. Currently, fondaparinux is indicated for the prophylaxis and treatment of venous thromboembolism.⁴³

Heparin also exhibits anti-inflammatory properties.⁴⁴ Although still to be fully clarified, some of the proposed mechanisms include binding with inflammatory cytokines, inhibition of neutrophil chemotaxis

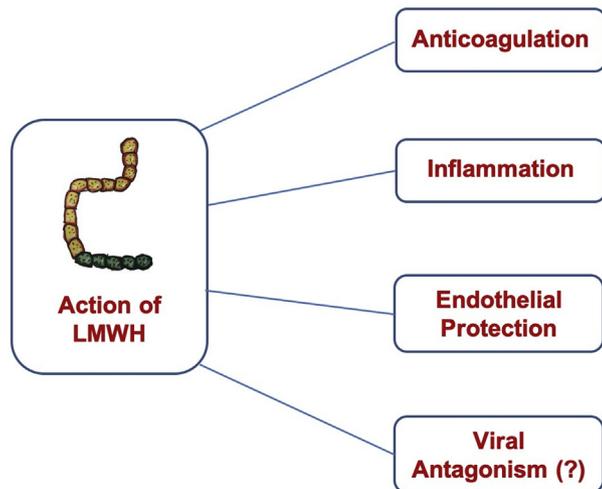


Fig 2. Potential effects of low-molecular-weight heparin (LMWH) in COVID-19 disease.

and leukocyte migration, neutralization of complement factor C5a and sequestration of acute phase proteins such as P-selectin and L-selectin, and induction of cell apoptosis through tumor necrosis factor α and nuclear factor κ B pathways.^{45,46} As described before, histones released from damaged cells can injure endothelial cells,⁴⁷ and heparin has been shown to antagonize histones and thus "protect" the endothelium.^{48,49} Another mechanism of heparin's action is characterized by its effects on histone methylation and on the mitogen-activated protein kinase and nuclear factor κ B signal pathways.⁵⁰ Therefore, heparin can protect from microcirculatory dysfunction and possibly reduce organ damage.

Experimental models put in light a potential antiviral role of heparin; indeed, its polyanionic polysaccharide strands can bind to different proteins, and therefore heparin could act as an effective inhibitor of viral adhesion.⁵¹ Competition between heparin and herpes simplex virus for host cell surface glycoproteins has been described, and in zika virus infection, heparin prevented virus-induced cell death of human neural progenitor cells.^{51,52} Furthermore, the addition of heparin to experimental

cultured cells halved the infection rate from sputum of a patient with severe acute respiratory syndrome pneumonia.⁵³

Surface plasmon resonance and circular dichroism, two spectroscopic techniques for molecular analysis, have been used in a recent work and showed that the binding domain of the Spike S1 SARS-CoV-2 protein receptor interacts with heparin.⁵⁴

Potential mechanisms of LMWH action in COVID-19 disease are described in Fig 2.

Finally, in the recent report by Tang et al,⁵⁵ a favorable outcome was highlighted with the use of LMWH in patients with severe COVID-19 disease meeting sepsis-induced coagulopathy criteria (Table) or with markedly high D-dimer levels. There were 99 patients who were treated with heparin for at least 7 days; in almost all patients (n = 94), enoxaparin was used subcutaneously (40-60 mg enoxaparin/d); 5 patients received unfractionated heparin (10,000-15,000 units/d).

CONSIDERATIONS AND POSSIBLE THERAPEUTIC IMPLICATIONS

Growing evidence puts emphasis on involvement of the coagulation system due to inflammation in COVID-19 pneumopathy. Previously, in H1N1 patients who developed acute respiratory distress syndrome, empirical systemic anticoagulation significantly reduced venous thromboembolism incidence without increased hemorrhagic complications.⁵⁶ Therefore, although data are still numerically insufficient to establish what the appropriate therapeutic regimen may be, the addition of heparin can have a favorable impact on progression of COVID-19 infection.

The reported data suggested that in many cases, the infection may have an asymptomatic course.⁵⁷ The use of heparin is probably unnecessary in this population. However, in case of the onset and persistence of respiratory symptoms, even in patients in isolation at home, it is considered useful to start prophylaxis with LMWH or with fondaparinux if renal function is preserved (creatinine clearance >50 mL/min). Should the patient develop a progressive worsening of respiratory symptoms in association with an increase in coagulation markers, therapeutic or subtherapeutic doses of

Table. Sepsis-induced coagulopathy (SIC) score

Category	Parameter	0 point	1 point	2 points
Prothrombin time	PT-INR	≤1.2	>1.2	>1.4
Coagulation	Platelet count ($\times 10^9/L$)	≥150	<150	<100
Total SOFA	SOFA four items ^a	0	1	≥2
Total score for SIC ≥4				

PT-INR, Prothrombin time-international normalized ratio; SOFA, sequential organ failure assessment.

^aTotal SOFA is the sum of the four items (respiratory SOFA, cardiovascular SOFA, hepatic SOFA, renal SOFA). The SOFA score was included to confirm the presence of sepsis but does not reflect sepsis severity; therefore, the score for SOFA was limited to 2 points even if the SOFA score was more than 2. (Adapted from Tang N, Bai H, Chen X, Gong J, Li D, Sun Z. Anticoagulant treatment is associated with decreased mortality in severe coronavirus disease 2019 patients with coagulopathy. *J Thromb Haemost* 2020;18:1094-9.)

LMWH could be hypothesized, taking into account the clinical characteristics and hemorrhagic risk; however, a definitive recommendation cannot be provided because of the lack of consistent published data. In advanced states, as powerful intravascular generation of thrombin occurs, unfractionated heparin could have a role, especially in cases of severe renal failure (creatinine clearance <30 mL/min) because of impaired LMWH renal excretion. Furthermore, in these patients, careful monitoring of the coagulation parameters is necessary because of the possible development of DIC in the end stage of the disease.⁴

The authors express their gratitude to Monica Grasso for her assistance in language editing.

AUTHOR CONTRIBUTIONS

Conception and design: LC

Analysis and interpretation: LC, FP, GA, PA, EA, GF

Data collection: LC, GF

Writing the article: LC

Critical revision of the article: LC, FP, GA, PA, EA, GF

Final approval of the article: LC, FP, GA, PA, EA, GF

Statistical analysis: Not applicable

Obtained funding: Not applicable

Overall responsibility: LC

REFERENCES

1. Klok FA, Kruij MJ, van der Meer NJ, Arbous MS, Gommers DA, Kant KM, et al. Incidence of thrombotic complications in critically ill ICU patients with COVID-19. *Thromb Res* 2020;191:145-7.
2. Danzi GB, Loffi M, Galeazzi G, Gherbesi E. Acute pulmonary embolism and COVID-19 pneumonia: a random association? *Eur Heart J* 2020;41:1858.
3. Xie Y, Wang X, Yang P, Zhang S. COVID-19 complicated by acute pulmonary embolism. *Radiol Cardiothorac Imaging* 2020;2:e200067.
4. Tang N, Li D, Wang X, Sun Z. Abnormal coagulation parameters are associated with poor prognosis in patients with novel coronavirus pneumonia. *J Thromb Haemost* 2020;18:844-7.
5. Goeijenbier M, van Wissen M, van de Weg C, Jong E, Gerdes VE, Meijers JC, et al. Viral infections and mechanisms of thrombosis and bleeding. *J Med Virol* 2012;84:1680-96.
6. Branchford BR, Carpenter SL. The role of inflammation in venous thromboembolism. *Front Pediatr* 2018;6:142.
7. Liao JK. Linking endothelial dysfunction with endothelial cell activation. *J Clin Invest* 2013;123:540-1.
8. Yun SH, Sim EH, Goh RY, Park JI, Han JY. Platelet activation: the mechanisms and potential biomarkers. *Biomed Res Int* 2016;2016:9060143.
9. Langer HF, Chavakis T. Leukocyte-endothelial interactions in inflammation. *J Cell Mol Med* 2009;13:1211-20.
10. Xu J, Lupu F, Esmon CT. Inflammation, innate immunity and blood coagulation. *Hamostaseologie* 2010;30:5-9.
11. Esmon CT, Xu J, Lupu F. Innate immunity and coagulation. *J Thromb Haemost* 2011;9:182-8.
12. Fuchs TA, Brill A, Wagner DD. Neutrophil extracellular trap (NET) impact on deep vein thrombosis. *Arterioscler Thromb Vasc Biol* 2012;32:1777-83.
13. Li J, Hara H, Wang Y, Esmon C, Cooper DK, Iwase H. Evidence for the important role of inflammation in xenotransplantation. *J Inflamm* 2019;16:10.
14. Robson SC, Shephard EG, Kirsch RE. Fibrin degradation product D-dimer induces the synthesis and release of biologically active IL-1 beta, IL-6 and plasminogen activator inhibitors from monocytes in vitro. *Br J Haematol* 1994;86:322-6.
15. Shorr AF, Thomas SJ, Alkins SA, Fitzpatrick TM, Ling GS. D-dimer correlates with proinflammatory cytokine levels and outcomes in critically ill patients. *Chest* 2002;121:1262-8.
16. Wu YP, Wei R, Liu ZH, Chen B, Lisman T, Ren DL, et al. Analysis of thrombotic factors in severe acute respiratory syndrome (SARS) patients. *Thromb Haemost* 2006;96:100-1.
17. Wong RS, Wu A, To KF, Lee N, Lam CW, Wong CK, et al. Haematological manifestations in patients with severe acute respiratory syndrome: retrospective analysis. *BMJ* 2003;326:1358-62.
18. Lee N, Hui D, Wu A, Chan P, Cameron P, Joynt GM, et al. A major outbreak of severe acute respiratory syndrome in Hong Kong. *N Engl J Med* 2004;348:1986-94.
19. Chong PY, Chui P, Ling AE, Franks TJ, Tai DY, Leo YS, et al. Analysis of deaths during the severe acute respiratory syndrome (SARS) epidemic in Singapore: challenges in determining a SARS diagnosis. *Arch Pathol Lab Med* 2004;128:195-204.
20. Hwang DM, Chamberlain DW, Poutanen SM, Low DE, Asa SL, Butany J. Pulmonary pathology of severe acute respiratory syndrome in Toronto. *Mod Pathol* 2005;18:1-10.
21. Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet* 2020;395:497-506.
22. Lippi G, Plebani M, Henry BM. Thrombocytopenia is associated with severe coronavirus disease 2019 (COVID-19) infections: a meta-analysis. *Clin Chim Acta* 2020;506:145-8.
23. Guan WJ, Ni ZY, Hu Y, Liang WH, Ou CQ, He JX, et al. Clinical characteristics of coronavirus disease 2019 in China. *N Engl J Med* 2020;382:1708-20.
24. Zhou F, Yu T, Du R, Fan G, Liu Y, Liu Z, et al. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. *Lancet* 2020;395:1054-62.
25. Levi M. Disseminated intravascular coagulation. *Crit Care Med* 2007;35:2191-5.
26. Smeeth L, Cook C, Thomas S, Hall AJ, Hubbard R, Vallance P. Risk of deep vein thrombosis and pulmonary embolism after acute infection in a community setting. *Lancet* 2006;367:1075-9.
27. Wiwanitkit V. Hemostatic disorders in bird flu infection. *Blood Coagul Fibrinolysis* 2008;19:5-6.
28. Ji HL, Zhao R, Matalon S, Matthay MA. Elevated plasmin(ogen) as a common risk factor for COVID-19 susceptibility. *Physiol Rev* 2020;100:1065-75.
29. Shodell M, Siegal FP. Circulating, interferon-producing plasmacytoid dendritic cells decline during human ageing. *Scand J Immunol* 2002;56:518-21.
30. Smits SL, de Lang A, van den Brand JM, Leijten LM, van Ijcken WF, Eijkemans MJ, et al. Exacerbated innate host response to SARS-CoV in aged non-human primates. *PLoS Pathog* 2010;6:e1000756.
31. Fulton RB, Varga SM. Effects of aging on the adaptive immune response to respiratory virus infections. *Aging Health* 2009;5:775.
32. Gralinski LE, Baric RS. Molecular pathology of emerging coronavirus infections. *J Pathol* 2015;235:185-95.

33. Yao XH, Li TY, He ZC, Ping YF, Liu HW, Yu SC, et al. A pathological report of three COVID-19 cases by minimally invasive autopsies. *Zhonghua Bing Li Xue Za Zhi* 2020;49:411-7.
34. Fox SE, Akmatbekov A, Harbert JL, Li G, Brown JQ, Heide RS. Pulmonary and cardiac pathology in COVID-19: the first autopsy series from New Orleans. *Lancet Respir Med* 2020 May 27. [Epub ahead of print].
35. Magro C, Mulvey JJ, Berlin D, Nuovo G, Salvatore S, Harp J, et al. Complement associated microvascular injury and thrombosis in the pathogenesis of severe COVID-19 infection: a report of five cases. *Transl Res* 2020;220:1-13.
36. Alquwaizani M, Buckley L, Adams C, Fanikos J. Anticoagulants: a review of the pharmacology, dosing, and complications. *Curr Emerg Hosp Med Rep* 2013;1:83-97.
37. Brinkhous K, Smith H, Warner E, Seegers W. The inhibition of blood clotting: an unidentified substance which acts in conjunction with heparin to prevent the conversion of prothrombin into thrombin. *Am J Physiol* 1939;125:683-7.
38. Lindahl U, Bäckström C, Höök M, Thunberg L, Fransson LA, Linker A. Structure of the antithrombin-binding site in heparin. *Proc Natl Acad Sci U S A* 1979;76:3198-202.
39. Lane DA, Denton J, Flynn AM, Thunberg L, Lindahl U. Anticoagulant activities of heparin oligosaccharides and their neutralization by platelet factor 4. *Biochem J* 1984;218:725-32.
40. Oosta GM, Gardner WT, Beeler DL, Rosenberg RD. Multiple functional domains of the heparin molecule. *Proc Natl Acad Sci U S A* 1981;78:829-33.
41. Hirsh J, Warkentin TE, Shaughnessy SG, Anand SS, Halperin JL, Raschke R, et al. Heparin and low-molecular-weight heparin: mechanisms of action, pharmacokinetics, dosing, monitoring, efficacy, and safety. *Chest* 2001;119:64S-94S.
42. Zhang Y, Zhang M, Tan L, Pan N, Zhang L. The clinical use of fondaparinux: a synthetic heparin pentasaccharide. *Prog Mol Biol Transl Sci* 2019;163:41-53.
43. Johnston A, Hsieh SC, Carrier M, Kelly SE, Bai Z, Skidmore B, et al. A systematic review of clinical practice guidelines on the use of low molecular weight heparin and fondaparinux for the treatment and prevention of venous thromboembolism: implications for research and policy decision-making. *PLoS One* 2018;13:e0207410.
44. Mousavi S, Moradi M, Khorshidahmad T, Motamedi M. Anti-inflammatory effects of heparin and its derivatives: a systematic review. *Adv Pharmacol Sci* 2015;2015:507151.
45. Oduah EI, Linhardt RJ, Sharfstein ST. Heparin: past, present, and future. *Pharmaceuticals (Basel)* 2016;9:38.
46. Thachil J. The versatile heparin in COVID-19. *J Thromb Haemost* 2020;18:1020-2.
47. Xu J, Zhang X, Pelayo R, Monestier M, Ammollo CT, Semeraro F, et al. Extracellular histones are major mediators of death in sepsis. *Nat Med* 2009;15:1318-21.
48. Iba T, Hashiguchi N, Nagaoka I, Tabe Y, Kadota K, Sato K. Heparins attenuated histone-mediated cytotoxicity in vitro and improved the survival in a rat model of histone-induced organ dysfunction. *Intensive Care Med Exp* 2015;3:36.
49. Zhu C, Liang Y, Li X, Chen N, Ma X. Unfractionated heparin attenuates histone-mediated cytotoxicity in vitro and prevents intestinal microcirculatory dysfunction in histone-infused rats. *J Trauma Acute Care Surg* 2019;87:614-22.
50. Ma J, Bai J. Protective effects of heparin on endothelial cells in sepsis. *Int J Clin Exp Med* 2015;8:5547-52.
51. Shukla D, Spear PG. Herpesviruses and heparan sulfate: an intimate relationship in aid of viral entry. *J Clin Invest* 2001;108:503-10.
52. Ghezzi S, Cooper L, Rubio A, Pagani I, Capobianchi MR, Ippolito G, et al. Heparin prevents Zika virus induced-cytopathic effects in human neural progenitor cells. *Antiviral Res* 2017;140:13-7.
53. Vicenzi E, Canducci F, Pinna D, Mancini N, Carletti S, Lazzarin A, et al. Coronaviridae and SARS-associated coronavirus strain HSR1. *Emerg Infect Dis* 2004;10:413-8.
54. Mycroft-West C, Su D, Elli S, Guimond S, Miller C, Turnbull J, et al. The 2019 coronavirus (SARS-CoV-2) surface protein (Spike) S1 receptor binding domain undergoes conformational change upon heparin binding. Available at: <https://www.biorxiv.org/content/10.1101/2020.02.29.971093v1.full>. Accessed June 26, 2020.
55. Tang N, Bai H, Chen X, Gong J, Li D, Sun Z. Anticoagulant treatment is associated with decreased mortality in severe coronavirus disease 2019 patients with coagulopathy. *J Thromb Haemost* 2020;18:1094-9.
56. Obi AT, Tignanelli CJ, Jacobs BN, Arya S, Park PK, Wakefield TW, et al. Empirical systemic anticoagulation is associated with decreased venous thromboembolism in critically ill influenza A H1N1 acute respiratory distress syndrome patients. *J Vasc Surg Venous Lymphat Disord* 2019;7:317-24.
57. Day M. Covid-19: four fifths of cases are asymptomatic, China figures indicate. *BMJ* 2020;369:m1375.

Submitted Apr 16, 2020; accepted May 22, 2020.